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LARGE-SCALE BIOLOGY

High-temporal-resolution Transcriptome Landscape of Early Maize Seed Development

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High-temporal-resolution transcriptomes uncover the genetic control of the developmental stages of double fertilization, coenocyte formation, cellularization and differentiation in early maize seed.

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ABSTRACT

The early maize (*Zea mays*) seed undergoes several developmental stages after double fertilization to become fully differentiated within a short period of time, but the genetic control of this highly dynamic and complex developmental processes remains largely unknown. Here, we report a high-temporal-resolution investigation of transcriptomes using 31 samples collected at an interval of 4 or 6 hours within the

first six days of seed development. These time-course transcriptomes were clearly separated into four distinct groups, corresponding to the stages of double fertilization, coenocyte formation, cellularization, and differentiation. A total of 22,790 expressed genes including 1,415 transcription factors (TFs) were detected in early stages of maize seed development. In particular, 1,093 genes including 110 TFs were specifically expressed in the seed and displayed high temporal specificity by expressing only in particular period of early seed development. There were 160, 22, 112 and 569 seed-specific genes predominantly expressed in the first 16 hours after pollination, coenocyte formation, cellularization and differentiation stage, respectively. In addition, network analysis predicted 31,256 interactions among 1,317 TFs and 14,540 genes. The high-temporal-resolution transcriptome atlas reported here provides an important resource for future functional study to unravel the genetic control of seed development.

INTRODUCTION

Maize (*Zea mays*) seed is one of the most important sources of food, feed and biofuel materials (Godfray et al., 2010), and serves as an excellent model for research on seed development due to its relatively large size. Maize seed development is initiated in the embryo sac with the fusion of the two pollen sperms with the egg cell and central cells of the female gametophyte to product the progenitors of embryo and endosperm, respectively (Chaudhury et al., 2001; Dumas and Mogensen, 1993). The embryo sac is embedded in the nucellus which will be gradually degraded after double fertilization. Nucellus degeneration is important for endosperm expansion and its products are believed to be taken up by endosperm (Greenwood et al., 2005; Russell, 1979). After double fertilization, the zygote undergoes an asymmetric division to form a small apical cell and a large basal cell, which develops into the embryo proper and the suspensor, respectively (Nardmann and Werr, 2009). The embryo proper further forms the mature embryo after the morphogenesis stage and will grow to be the next plant generation (Nardmann and Werr, 2009). The development of endosperm begins with the formation of a coenocyte, which the primary endosperm undergoes several

rounds of nuclear divisions but without cytokinesis. The coenocyte then undergoes cellularization and cell differentiation (Olsen, 2001; Leroux et al., 2014; Lopes and Larkins, 1993; Sabelli and Larkins, 2009). After differentiation, the endosperm enlarges significantly through further cell division, cell expansion and endoreduplication. Different from dicots which the endosperm is mostly consumed or absorbed by the developing embryo, maize endosperm serves as a storage tissue to store the proteins and carbohydrates needed for seedling development (Berger, 1999; Olsen, 2001; Lopes and Larkins, 1993; Sabelli and Larkins, 2009).

Understanding the spatial and temporal gene expressional profile along seed development is helpful for the genetic improvement of this important crop. Over the years, several transcriptome profiling studies have been conducted to detect the expressed genes and cellular processes for seed development in *Arabidopsis thaliana* (Belmonte et al., 2013; Le et al., 2010), rice (Gao et al., 2013; Xu et al., 2012), *Tropaeolum majus* (Jensen et al., 2012), and soybean (*Glycine max*; Jones and Vodkin, 2013). In maize, the transcriptome of endosperm was initially characterized using expressional sequence tag (EST) sequencing method (Lai et al., 2004). The dynamic of gene expression during seed development was then investigated by microarray-based approach, which identified 3,445 genes with differential expression among samples of six different time points (Liu et al., 2008). The general transcriptome-wide differences between embryo and endosperm had also been analyzed in maize seed of 9 day after pollination (DAP) using RNA-seq method (Lu et al., 2013). Then more detail transcriptome atlas of maize seed development were generated using RNA-seq data from embryo, endosperm, and intact seed sampled at an interval of two days from 0~38 DAP, which providing an extensive view of transcriptome dynamics over seed development (Chen et al., 2014). To gain the information of spatial distribution of genes in endosperm, a laser-capture microdissection (LCM) study was reported at 8 DAP, which allowed the identification of a number of compartment specifically expressed genes in the endosperm of this particularly stage (Zhan et al., 2015). Recently, the transcriptomes of isolated mature

female and male gametes, 12 and 24 hours after pollination (HAP) zygote, and apical and basal daughter cells were also obtained (Chen et al., 2017).

In maize, the double fertilization events typically finish in the first DAP (Sabelli and Larkins, 2009), with an average of 8 HAP (Chen et al., 2017). The coenocytic stage of maize endosperm usually occurs during 1 to 2 DAP, and then is followed by a period of cellularization at about 3 to 4 DAP (Leroux et al., 2014; Sabelli and Larkins, 2009). The endosperm cell differentiation starts at about 5 DAP, forming four main cell types: starchy endosperm (SE), aleurone (AL), embryo-surrounding region (ESR), and basal endosperm transfer layer (BETL) (Leroux et al., 2014; Olsen, 2001; Sabelli and Larkins, 2009). In line with the rapid transition of these developmental stages, large numbers of genes are involved in the key steps of double fertilization, coenocyte formation, cellularization and differentiation that happen in the first few days of seed development, but these genes may not have been captured in the above-mentioned extensive transcriptome studies. For instance, *embryo sac1* (*ES1*) and *embryo sac4* (*ES4*), two genes encoding secreted peptides and required for micropylar pollen tube guidance and burst, are only expressed in the nucellus during the first few HAP and then show low or even no expression a few hours later (Chen et al., 2017; Cordts et al., 2001). Therefore, it is highly possible that many genes that are important for the early seed development, but are expressed only in a short period of time or particular developmental stages have not been identified yet, due to the fact that the previous transcriptome studies did not have sufficient temporal resolution.

Here we report a comprehensive high-temporal-resolution investigation of transcriptomes using data for 31 time points, at 4- or 6-hour intervals within the first six days of maize seed development. This high-density time-course transcriptome analysis clearly highlighted the timings of double fertilization, coenocyte formation, cellularization and differentiation in the endosperm. In total, 22,790 genes, including 1,415 transcription factors (TFs), were found to be expressed during early maize seed development. These genes were classed into 18 coexpression modules according their

expression patterns, which provided further insight into the dynamics of transcriptome reprogramming underlying the developmental and physiological transitions of the four distinct development stages. A total of 1,093 genes, including 110 TFs, specifically expressed in seed were identified and most of these seed-specific genes had high temporal specificity, being expressed only in a particular period of time within the first six days of maize seed development. TF regulatory network analysis predicted 31,256 interactions among 1,317 TFs and 14,540 seed-expressed genes. The high-temporal-resolution transcriptomes presented here provide a valuable resource for the study of seed biology.

RESULTS

The generation of high-temporal-resolution transcriptome data at early stages of maize seed development

To investigate gene activity dynamic during early maize seed development, we carried out RNA-seq for the nucellus (embryo sac included) of inbred line B73 from 0~144 HAP with an interval of 4 hours (0~72 HAP) or 6 hours (72~144 HAP) (**Figure 1** and **Supplemental Figure 1**). Two biological replicates, which each were pooled samples from at least three plants, were set up for all 31 time points. Totally, 2.85 billion high-quality reads were generated using Illumina sequencing platform, and then mapped to the maize B73 reference genome (RefGen_V4) (Jiao et al., 2017) using Hisat (Kim et al., 2015). An average about 93% of reads were uniquely mapped (**Supplemental Table 1**) and only the uniquely mapped reads were further used to calculate normalized gene expression level as FPKM (fragments per kilobase of transcript per million mapped reads). Comparison of the two biological replicates showed that the expression values between them were highly correlated (average R^2 = 0.94). Hence, we took the average FPKM value of the two replicates as expression level for the sample at each time point. To reduce the influence of transcription noise, here we defined a gene as expressed if its FPKM value was ≥ 1. In total, 22,790 genes including 1,415 TFs were found to be expressed in at least one of the 31 samples

(**Supplemental Data Set 1 and Supplemental Data Set 2**).

To further validate the quality of the gene activity profiles obtained here, we specifically examined the expression patterns of 8 genes for which transcript levels were previously reported during early maize seed development. *ZmMCM3*, *ZmMCM6*, *ZmCYC1*, and *ZmCYC3* are genes involved in the cell cycle process and were shown to be induced after fertilization (Dresselhaus et al., 1999; Dresselhaus et al., 2016; Sauter et al., 1998). The expression of *ZmMCM3*, *ZmMCM6*, *ZmCYC1*, and *ZmCYC3* are induced in the zygote at 12 and 24 HAP, and *ZmCYC1* and *ZmCYC6* reached highest expression later than that of *ZmMCM3* and *ZmMCM6* (Chen et al., 2017). Here, we found the expression of these four genes began to increase at 8 HAP, *ZmMCM3* and *ZmMCM6* showed the highest expression around 20 HAP, and *ZmCYC1* and *ZmCYC3* showed the highest expression around 32 HAP (**Supplemental Figure 2**). In addition, *Esr2*, a gene specially expressed in ESR (Bonello et al., 2000), and *Betl10*, a gene related to the differentiation of BETL (Zhan et al., 2015), were expressed after 102 HAP (**Supplemental Figure 2**), consistent with the idea that endosperm differentiation usually happened at about 4~6 DAP (Sabelli and Larkins, 2009). We also found that *ZmSWEET4C*, a hexose transporter gene predominantly expressed in BETL (Sosso et al., 2015), was highly expressed after 102 HAP and that *ZmYUC1*, an auxin biosynthesis gene (Bernardi et al., 2012; Doll et al., 2017), was rapidly activated after 126 HAP (**Supplemental Figure 2**), similar to their expression patterns reported previously (Doll et al., 2017; Li et al., 2014). In summary, the expression dynamics of these genes are in line with previous reports, indicating high quality and reliability of our data.

High-temporal-resolution transcriptomes can be clustered into four groups corresponding to different developmental stages

To gain insight into the transcriptome dynamic of early maize seed development, we performed hierarchical clustering (**Figure 2A**) and principal component analysis (PCA) (**Figure 2B**) for the 31 time-series samples. In line with the previously

reported timing of double fertilization, coenocyte formation, cellularization and differentiation stages for early maize seed development (Chen et al., 2017; Sabelli and Larkins, 2009; Leroux et al., 2014; Olsen, 2001), these high-density time series transcriptomes can be generally divided into four groups with each group corresponding to a specific developmental stage (**Figure 2C**).

The samples from earliest time points (0~16 HAP) formed the first cluster and represented the stage around double fertilization (Stage I). *WAX2* encodes secreted peptides relating to pollen fertility as reported in Arabidopsis and cucumber (*Cucumis sativus*; Chen et al., 2003; Wang et al., 2015). G*lutamate decarboxylase protein* (*GAD*) encodes a non-protein amino acid that plays an important role in pollen tube growth and guidance (Akama and Takaiwa, 2007; Jin et al., 2016). In line with the process of double fertilization, we found both *ZmWAX2* and *ZmGAD* were highly expressed at this stage but were low or even not expressed in later time points (**Figure 2D**). It was reported that heat shock protein (HSP) and heat shock transcription factor (HSF) are involved in the regulation of reproductive system development, germ cell development and fertilization in mouse and human (Le Masson et al., 2011; Nixon et al., 2017). Here we found that *ZmHSP20* and *ZmHSF24* displayed increased expression after 8 HAP, but rapidly decreased after 16 HAP (**Figure 2D**), which suggested that *ZmHSP20* and *ZmHSF24* might be important around fertilization in maize.

The samples between time points 20 HAP and 44 HAP formed a second cluster and represented the stage of coenocyte formation in endosperm (Stage II). In this stage, the initial triploid nucleus undergoes several rounds of synchronous division in the absence of cell wall formation and cytokinesis, resulting in the formation of a coenocytic endosperm. As reported in many organisms, canonical H3 genes are expressed during S-stage of the cell cycle and are DNA replication dependent (Ahmad and Henikoff, 2002; Cui et al., 2006; Hamiche and Shuaib, 2013; Otero et al., 2014). According to highly active DNA replication at coenocytic stage, two canonical H3

genes, *ZmH3-a* (Zm00001d042730) and *ZmH3-b* (Zm00001d045268) showed predominant expression at coenocytic stage in maize (**Figure 2E**). *WRKY10* in Arabidopsis is a regulator of seed size and is expressed in the developing endosperm from the two-nuclei stage at ~12 hours post fertilization to endosperm cellularization at ~96 hour (Luo et al., 2005). Here we found its homologous genes in maize, *ZmWRKY53* and *ZmWRKY104*, were mainly expressed at coenocytic stage (**Figure 2E**), which suggested that *ZmWRKY53* and *ZmWRKY104* might be important for endosperm proliferation in maize.

Samples between 48 HAP to 96 HAP fell into the third cluster, which correspond to the cellularization stage (Stage III). *ZmExo1* (Zm00001d017799) encodes an RNA exonuclease. The RNA exonuclease is required for mitotic cell division in *Schizosaccharomyces* (Snee et al., 2016). Collaborative control of cell cycle progression by the RNA exonuclease protein is conserved across species (Snee et al., 2016). *ZmLac* (Zm00001d018601) encodes a laccase that contributes to cell-wall reconstitution in regenerating protoplasts of higher plants (Mayer and Staples, 2002). As reported in rice, the laccase gene *OsLac* could affect grain yield (Mayer and Staples, 2002). *ZmTOM* (Zm00001d040440) encodes a translocase of the outer mitochondrial membrane (TOM) which can transport mitochondrial precursor proteins (Wiedemann et al., 2003). Previous reports showed that TOM plays an important role in regulation of the cell cycle (Harbauer et al., 2014; Westermann, 2010). *ZmGCRP* (Zm00001d028862) encodes a glycine and cysteine rich family protein precursor (GCRP). The GCRP proteins play crucial roles in cell–cell signaling and participate in cell division and proliferation in rice (Harbauer et al., 2014; Westermann, 2010). Consistent with the active cell division and cell wall formation that occurs during the cellularization stage, we found these four genes were mainly expressed at this period (**Figure 2F**).

The fourth cluster was from 102 HAP to 144 HAP, which corresponded to the initial stage of differentiation in the endosperm (Stage IV). *Esr1* is an endosperm-specific

gene expressed in a restricted region around embryo and might involve in the establishment of physical barrier between embryo and endosperm (Harbauer et al., 2014; Westermann, 2010). *MYB-related protein 1* (*MRP1*) and *Betl3* are two BETL-specific genes important for the development and differentiation of BETL (Gomez et al., 2009; Hueros et al., 1999; Zhan et al., 2015). *Al9* is a gene related to AL differentiation (Gomez et al., 2009). We found all these four genes showed a rapidly increased expression at stage IV (**Figure 2G**), indicating that this stage is typically by the initiation of endosperm differentiation. In summary, our results demonstrated that our high-temporal-resolution transcriptome data are powerful for the stage specific genes, and that the dynamic transcriptome during the early endosperm development can be separated into four distinct groups corresponding to four different developmental stages.

Gene expression at different developmental stages of early maize seed

The global hierarchical clustering and PCA analysis graphically display the four main developmental stages of early maize seed. To further provide insights into the functional transitions along early seed development, we clustered all 22,790 expressed genes, including 1,415 (6.2%) TFs (**Supplemental Data Set 1 and Supplemental Data Set 2**), into 18 coexpression modules using the k-means clustering algorithm, and then performed MapMan annotation to assign genes to functional categories for each module (**Figure 3 and Supplemental Figure 3)**. Of which, genes that belong to the first nine modules were mainly expressed at only one of the four developmental stages and represented the particular functions for their corresponding stages (**Figure 3A**).

Genes expressed around double fertilization (Stage I) The stage around double fertilization (0~16 HAP) is best represented by 4,453 expressed genes, including 414 TFs, in modules I-A to I-D (**Figure 3A and Supplemental Data Set 1**). The module I-A (535 genes, 51 TFs) contains a set of genes related to protein serine/threonine kinase activity and amino acid phosphorylation (**Figure 3B**). These genes might

involve in the initial pollination response as they mainly expressed at 0 to 4 HAP. The module I-B (1,718 genes, 136 TFs) corresponds to genes which were highly expressed at 0 to 12 HAP and then appeared to be low or not expressed at later time points (**Figure 3A**). As reported previously, fertilization occurs at about 8 HAP on average (Chen et al., 2017). During fertilization, the pollen tube extends to the embryo sac to release sperm to form the zygote (Faure et al., 2003; Luo et al., 2005). Zygotes need energy (ATP) and thus produce energy-rich metabolites for generating ATP (Labarca and Loewus, 1973; Obermeyer et al., 2013; Rounds et al., 2011). Therefore, the genes in module I-B might contribute to the growth of pollen tube as they were overrepresented by genes involved in ATP binding, helicase activity, nucleotide binding and nucleoside triphosphatase activity (**Figure 3B**). For example, *ES1* and *ES4* in module I-B (**Supplemental Data Set 1**) relate to the pollen tube growth arrest and burst (Cordts et al., 2001).

Ca^{2+} signaling is thought to play important roles in plant growth and development, including key aspects of pollen tube growth and fertilization (Dresselhaus et al., 2016; Schiott et al., 2004). In module I-B, we found there were 17 genes involved in the calcium signaling pathway (**Supplemental Data Set 1**), including Zm00001d031543. Its homolog in Arabidopsis, *CA^{2+}-ATPASES 9* (*ACA9*), functions in the pollen tube plasma membrane and is a key regulator of pollen tube growth and fertilization (Schiott et al., 2004).

The genes in module I-C (1,463 genes,122 TFs) were highly expressed at 8~16 HAP and represented by genes related to transcription regulation (**Figure 3B**). MYB TFs are involved in controlling various processes like responses to abiotic and biotic stresses, differentiation, development, metabolism, defense (Ambawat et al., 2013; Yanhui et al., 2006). We found seven MYB TFs (*MYB3*, *MYB32*, *MYB38*, *MYB81*, *MYB112*, *MYB126* and *MYB133*) and five MYB related TFs (*MYBR26*, *MYBR51*, *MYBR78*, *MYBR84* and *MYBR90*) in module I-C, reflecting the important role of MYB TFs in early seed development. Moreover, nine important plant-specific GARS

TFs (*GRAS3*, *GRAS27*, *GRAS34*, *GRAS39*, *GRAS50*, *GRAS61*, *GRAS82*, *GRAS83* and *GRAS84*) and ten ethylene responsive AP2-EREBP TFs (*EREB8*, *EREB96*, *EREB117*, *EREB131*, *EREB156*, *EREB158*, *EREB159*, *EREB162*, *EREB192* and *EREB201*) were also found in module I-C.

Interestingly, in module I-D, there were 737 genes, including 105 TFs, only highly expressed at about 12 HAP we analyzed (**Figure 3A**). To further confirm the expression patterns of genes in module I-D, we analyzed the published RNA-seq data of isolated maize gametes, zygotes, and apical and basal daughter cells (Chen et al., 2017). For 512 genes detected in Chen's samples (Chen et al., 2017), about 75% of which were induced in zygote formation, with high expression at 12 HAP but low expression at 24 HAP (**Supplemental Figure 4 and Supplemental Data Set 3**), indicating the genes in module I-D we identified were indeed transitorily activated after fertilization. The genes in module I-D were enriched in TF activity, transcription regulation and sequence-specific DNA-binding (**Figure 3B**), suggesting most of which might be key regulatory function at the beginning of new generation formation. For example, *lipoxygenase-1* (*LOX1*) in module I-D is important for regulation of defense-related signaling molecules and activation of the antioxidative enzyme system (Cho et al., 2012). We also found there were three mitogen-activated protein kinase (MPKs) genes, *MPK1*, *MPKKK11* and *MPKKK18*, in module I-D (**Supplemental Data Set 1**). It was showed that MPK cascades function as molecular switches in response to spatiotemporal specific ligand–receptor interactions and the availability of downstream substrates, and are ubiquitous signaling modules in eukaryotes (Group, 2002; Widmann et al., 1999; Xu and Zhang, 2015).

Genes expressed during the coenocyte formation stage (Stage II). The stage of coenocyte formation (20~44 HAP) is best represented by 1,285 genes, including 53 TFs, in module II (**Figure 3A and Supplemental Data Set 1**). Consistent with the active chromatin formation and nuclear division that occurs at the coenocytic stage, module II was overrepresented by genes related to DNA replication, transcription

initiation factor activity, microtubule-based movement, microtubule motor activity, nucleosome assembly, nucleosome, transcription initiation, DNA replication initiation and DNA binding (**Figure 3B**). Histones, the major protein components of chromatin, are highly conserved amongst eukaryotes (Ingouff and Berger, 2010). Based on the reported histone sequences in Arabidopsis (Ingouff and Berger, 2010; Kawashima et al., 2015; Okada et al., 2005; Ramesh Yelagandula, 2014), we identified a total of 79 histone genes in the maize genome, and found 66 of these were expressed in our data, including 6 *H1*, 4 *H2A*, 6 *H2A.W*, 3 *H2A.X*, 5 *H2A.Z*, 13 *H2B*, 12 *H3*, 5 *H3.3* and 12 *H4* (**Supplemental Data Set 4)**. Of these 66 expressed histone genes, 71% (47) belonged to module II, including 3 *H1*, 2 *H2A*, 5 *H2A.W*, 1 *H2A.X*, 1 *H2A.Z*, 12 *H2B*, 12 *H3* and 11 *H4* (**Figure 4 and Supplemental Data Set 4**). Notably, all 12 expressed *H3* were in module II with predominant expression during the coenocyte stage, and all 5 expressed *H3.3* did not show up in module II. This result was consistent with the reports that canonical H3 deposition is coupled to DNA synthesis during replication and repair, which is extremely activated for coenocyte formation, whereas H3.3 is deposited independently of replication (Ahmad and Henikoff, 2002; Cui et al., 2006). A number of previous works showed that different histone H2A variants have distinct functions in diverse biological processes (Kawashima et al., 2015; Talbert and Henikoff, 2010, 2014; Weber and Henikoff, 2014). H2A.W is specific to seed-bearing plants and predominantly localizes in heterochromatin to promote heterochromatin condensation (Ramesh Yelagandula, 2014). We found five of six expressed *H2A.W* genes were in module II. By contrast, for 5 expressed *H2A.Z*, only one *H2A.Z* (Zm00001d027760) was in module II, and the remained 4 *H2A.Z* were distributed in four different modules (I-C,11,16,18) (**Supplemental Data Set 4**). The high variation of expression patterns for different *H2A.Z* genes is line with the diverse functions of H2A.Z variants, including DNA repair, apparently contradictory roles in gene activation and silencing, nucleosome turnover, heterochromatin, boundary element and chromatin fiber formation (Altaf et al., 2009; Dai et al., 2017; Domaschenz et al., 2017; Zlatanova and Thakar, 2008).

***Genes expressed during the cellularization stage (Stage III*).** The stage of endosperm cellularization (48~96 HAP) is best represented by 2,569 expressed genes, including 125 TFs, in modules III-A and III-B (**Figure 3A and Supplemental Data Set 1**). The genes in module III-A (1,343 genes, 73 TFs) were highly expressed during the entire cellularization stage (48~96 HAP) (**Figure 3A**). We found there were 14 auxin pathway genes (Hagen and Guilfoyle, 2002; Chen et al., 2017; Yue et al., 2015) in module III-A, including three ATP-Binding Cassette Subfamily B Members (ZmABCBs: *ZmABCB11*, *ZmABCB30* and *ZmABCB35*) representing potential auxin transporter genes, three auxin-responsive factors (ZmARFs: *ZmARF2*, *ZmARF7* and *ZmARF15*), four genes encoding proteins that interact with ARF regulators (ZmIAAs: *ZmIAA2*, *ZmIAA7*, *ZmIAA15* and *ZmIAA23*), and four auxin response genes (ZmSAURs (Small auxin up RNAs): *ZmSAUR4*, *ZmSAUR22*, *ZmSAUR31* and *ZmSAUR56*). These results reflected the importance of auxin transporter and response genes in endosperm cellularization. The genes in module III-B (1,226 genes, 52 TFs) were mainly expressed at late of cellularization stage (72~96 HAP). Genes related to membrane and protein binding were enriched in III-B, which might be involved in the formation of cell membrane during cellularization (**Figure 3B**). For example, syntaxins are membrane proteins involved in vesicle trafficking and release of neurotransmitters (Besserer et al., 2012; Burgess et al., 1997; Jung et al., 2012). In maize, the syntaxin protein SYP121 could selectively regulate plasma membrane aquaporin trafficking (Besserer et al., 2012). Here we found four syntaxin genes, which we named *ZmSYP121a* (Zm00001d020187), *ZmSYP121b* (Zm00001d041716), *ZmSYP121c* (Zm00001d042018) and *ZmSYP121d* (Zm00001d048147), were expressed in stage III-B, suggesting they might be involved in cell membrane formation during cellularization.

Genes expressed during the differentiation stage (Stage IV). The stage of initial endosperm differentiation (102~144 HAP) is best represented by 3,614 genes, including 224 TFs, in module IV-A and IV-B (**Figure 3A and Supplemental Data Set 1**). The genes in module IV-A (1,357 genes, 81 TFs) were highly expressed from

about 102 HAP (**Figure 3A**), while genes in module IV-B (2,257 genes, 143 TFs) were highly expressed from about 114 HAP. These genes might be mainly involved in differentiation. Many genes specifically expressed in endosperm subregions were included in these two modules, including two AL marker genes, *VPP1* (IV-A) (Wisniewski and Rogowsky, 2004) and *Al9* (IV-B) (Gomez et al., 2009); four ESR specific genes, *Esr1* (IV-B), *Esr2* (IV-B), *Esr3* (IV-B) and *Esr6* (IV-B) (Balandin et al., 2005; Bonello et al., 2000; Magnard et al., 2000; Opsahl-Ferstad et al., 1997; Todorow et al., 2018; Zhan et al., 2015); and 8 BETL specific genes, *Ebe2* (IV-A), *MYBR33* (IV-B), *MRP1* (IV-B), *Betl-3, 9, 10* (IV-B), *Bap2* (IV-B) and *Mn1* (IV-B) (Cheng et al., 1996; Gomez et al., 2009; Hueros et al., 1999; Magnard et al., 2003; Serna et al., 2001; Zhan et al., 2015).

Genes expressed during more than one of the four stages. We found a total of 10,869 genes, including 599 TFs, in modules of M10~M18 were expressed at more than one of the four stages (**Figure 3A**), indicating there are some common functional processes in different stages. For example, genes related to microtubule associated complexes were enriched in M10 (**Supplemental Figure 3**). They displayed continuous expression at fertilization and coenocytic stages (0~44 HAP), which is consistent with the report that microtubule associated complexes are involved in fertilization, mitosis and cell division (Schatten et al., 1985). In line with the observation that photosynthetic genes are expressed during the development of seed in Arabidopsis (Schmid et al., 2005), here we found genes related to chlorophyll biosynthetic processes were overrepresented in M10 (**Supplemental Figure 3**), such as the *Chlorophyll synthase G1* (*CHLG1*) (Hunter et al., 2018) and magnesium chelatase gene *Oil yellow 1* (*Oy1*) (Sawers et al., 2006), both required for chlorophyll biosynthesis. In addition, we found *Geranylgeranyl hydrogenase 1* (*Ggh1*) (Owens et al., 2014), the homolog of rice *Geranylgeranyl reductase* (*OsCHL P*) for chlorophyll synthesis (Wang et al., 2014), was also included in M10. These implied that the photosynthesis system might start to be established in early development seed in maize. Maize is generally considered as a chilling sensitive species (Miedema, 1982).

We found genes related to homoiothermy, ice binding and response to freezing are enriched in M13, with continuous expression at fertilization, coenocytic and cellularization stages (0~96 HAP) (**Figure 3A and Supplemental Figure 3**). This suggested these cold-response genes might function to stabilize membranes against freeze-induced injury and help seed to develop under suboptimal temperature conditions.

In addition, as shown in M16, 1,338 genes including 27 TFs were activated after double fertilization, with continuous expression at stages II~IV. These genes were enriched in many basic functional categories processes, including structural constituent of ribosome, cytoplasm, translation, intracellular, translational elongation, small ribosomal subunit, GTPase activity, RNA binding, protein catabolic process and vesicle-mediated transport.

Seed-specific genes including TFs and their target genes involved in the four stages of early maize seed development

The high-temporal-resolution transcriptome profiling data generated here provided us a good opportunity to identify genes specifically expressed in particular stages of early seed development, which is highly informative for inferring gene function and understanding the genetic control of developmental transition. Combined with 23 published non-seed transcriptome data sets, including root, shoot, shoot apical meristem, leaf, cob, tassel and immature ear (Bolduc et al., 2012; Davidson et al., 2011; Jia et al., 2009; Li et al., 2010; Wang et al., 2009), we identified a total of 1,093 genes specifically expressed in seed (**Supplemental Figure 5 and Supplemental Data Set 5**). Of these, 654 genes were not found in our previous transcriptomic study in maize seed (Chen et al., 2014) and in a recent study based on the extensive transcriptomes of 79 B73 tissues (Hoopes et al., 2018). This is likely because for the first 6 DAP of maize seed development, the intact seed samples of only 5 and 3 time points were analyzed by Chen et al. (Chen et al., 2014) and Hoopes et al. (Hoopes et al., 2018), respectively. However, the transcriptome data of 31 time points generated

in this study enabled us to identify additional early seed-specific genes, especially those specifically expressed in a short period of time. The seed-specific genes identified here accounted for about 6.20% of all expressed genes detected. By contrast, for 1,415 TFs detected as expression in our data, 10.1% (110) of which are seed-specific TFs (**Supplemental Data Set 5**). The seed-specific genes and TFs were significantly enriched in the stage of differentiation (after 102 HAP) (**Supplemental Figure 6**), reflected the important role of seed-specific genes and TFs for the generation of new tissue or cell types during differentiation.

We inferred the gene regulatory network (GRN) that connects TFs with their potential target genes using the method reported previously (Faith et al., 2007; Xiong et al., 2017). The GRN is scale free, based on the frequency distribution of the degree of the nodes (**Supplemental Figure 7**). In total, 14,540 target genes (including 1,317 TFs) were included in our newly constructed GRN with a total of 31,256 interactions (**Supplemental Data Set 7**). Next, we focused on the network communities connected with seed-specific TFs. In total, 81 seed-specific TFs and 1,483 potential target genes were identified, with a total of 2,000 interactions. As expected, seed-specific genes were more likely to be regulated by seed-specific TFs as compared with non-seed-specific genes. Among 513 seed-specific genes in the GRN, 40% (205) of which displayed a total of 407 interactions with 57 seed-specific TFs. By contrast, for 13,675 non-seed-specific genes in the GRN, only 9.3% (1,278) of which were interacted with 79 seed-specific TFs, resulting a total of 1,593 interactions.

Next, combined with the transcriptome data generated previously (Chen et al., 2014), we further explored the expression patterns of 1,093 seed-specific genes we identified in later development stage of embryo and endosperm (**Figure 5 and Table 1)**.

Specific genes expressed around double fertilization (Stage I). 160 seed-specific genes, including 18 TFs, in group I were mainly expressed around double fertilization

(0~16 HAP) (**Figure 5 and Supplemental Data Set 5**), and in this group, genes related to transcription factor activity, enzyme inhibitor activity, sequence-specific DNA binding, pectin esterase activity and hydrolase activity are overrepresented **(Supplemental Figure 8**). The high expression of genes related to enzyme inhibitor activity at fertilization stage might be associated with the protection of female reproductive cells from a variety of biotic stresses, including enzymes from nucellus or synergid lysate, contents of burst pollen tubes and pathogen attack (McInnis et al., 2006). For example, *ES1* and *ES4*, encoding peptides with structural homology to defensins (proteinase inhibitors) in group I, are expressed in the embryo sac and were suppressed after fertilization (Cordts et al., 2001).

We found 13 seed-specific TFs in group I were included in the GRN, with a total of 148 interactions. The top 5 seed-specific TFs with the most connections in group I are *HSF24*, *HSF20*, *EREB117*, *BZIP29* and *GRAS61* (**Supplemental Figure 9 and Supplemental Data Set 8**). *HSF24* and *HSF20* were predicted to interact with 32 and 24 genes, respectively. Notably, *HSF24* and *HSF20* were also highly expressed and ranked as the first and 10th highly expressed seed-specific genes in stage I (0~16 HAP) (**Supplemental Data Set 5**). In mice, HSF1 is the major regulator for heat shock transcriptional response, and HSF1-deficient mice exhibited complex phenotypes, including developmental defects and complete female infertility (Le Masson et al., 2011). Here, our results suggested that HSF24 and HSF20 might be the major regulators of heat shock transcriptional response and play important roles around double fertilization in maize seed. The AP2 family TF gene *EREB117* was predicted to interact with 20 genes. Its homolog *WRI1* in Arabidopsis is involved in the control of storage compound biosynthesis, which mutant has wrinkled seed phenotype (Hanano et al., 2018). *BZIP29* was predicted to interact with 13 genes, including three known genes (*EMP10, RH4* and *Pco090181a*) and two known TFs (*GRAS61* and *C2H2*). Of these, the *EMP10* encodes a mitochondrial PPR protein and is required for embryogenesis and endosperm development in maize (Cai et al., 2017).

GRAS are plant-specific TFs and play important roles in many processes such as signal transduction, stress responses and meristem maintenance (Bolle, 2004; Mayrose et al., 2006; Zhang et al., 2018). At present, only a few GRAS proteins have been characterized in maize, such as ZmGRAS20, which was specifically expressed in endosperm and involved in regulating starch biosynthesis (Cai et al., 2017). Here, our results showed that *GRAS61* was mainly expressed around double fertilization and was predicted to interact with 11 genes, including *ES4*, an embryo sac specific gene playing an important role in fertilization process (Cordts et al., 2001).

Specific genes expressed during coenocyte formation stage (Stage II). Group II contains 22 seed-specific genes mainly expressed during coenocyte formation (20~44 HAP) (**Figure 5 and Supplemental Data Set 5**). *BURP domain-containing protein-RD22-like9* (*BURP9*) was included in this group. The BURP domain-containing gene family is a large plant-specific gene family, yet their functions are very poorly understood, especially in maize. *BURP9* was reported to respond to ABA and cold (Gan et al., 2011). Here, we found *BURP9* was mainly expressed during coenocyte formation, which will be helpful for further understanding its function in maize.

ZmCoe4, the only seed-specific TF with predominant expression during coenocyte formation, encodes a WRKY family TF (**Supplemental Figure 9 and Supplemental Data Set 8**). *ZmCoe4* was predicted to interact with 16 genes, including *15 kDa zein protein* (*ZP15*), *pathogenesis-related protein3* (*PRP3*), *glucan endo-1,3-beta-glucosidase homolog1* (*GEB1*), *chitinase A1* (*CTA1*), *umc2348* and *wound induced protein1* (*WIP1*) (Supplemental Data Set 8).

Specific genes expressed during cellularization stage (Stage III). Group III represents 112 seed-specific genes, including 7 TFs, predominantly expressed at the cellularization stage (48~96 HAP) (**Figure 5 and Supplemental Data Set 5**). *Zea AGAMOUS homolog 2* (*ZAG2*) was included in this group. *ZAG2*, which is

548 homologous to the Arabidopsis floral homeotic gene *AGAMOUS*, is expressed in
549 developing ovules and the inner carpel faces and might be important for maize flower
550 development (Schmidt et al., 1993). Here we found *ZAG2* was highly expressed after
551 pollination, especially at the cellularization stage (48-96 HAP), which is consistent
552 with the report in *Orchis italica* that *OitaAG* mRNA levels were high in columns and
553 ovaries (Salemme et al., 2013), particularly after pollination. However, up to now no
554 function has yet been determined for ZAG2.
555
556 The top 5 seed-specific TFs with the most connections in group III are *MYB131*,
557 *MYB16*, *BZIP109*, *ZAG2* and *BZIP114* (**Figure 6A, Supplemental Figure 9 and**
558 **Supplemental Data Set 8**). *MYB131*, *MYB16* and *BZIP109* were predicted to interact
559 with 109, 83 and 51 genes, respectively, which some of their potential target genes are
560 overlapped (**Figure 6A**). There were 18 genes including three known TFs (*MYB8*,
561 *Knox1* and *GGATA12*) that were predicted to be regulated by MYB16 and BZIP109,
562 and there were 17 genes including four previous characterized genes (*Sumo1a*,
563 *Cdpk13*, *Bm1* and *AY107053*) were predicted to be regulated by *MYB16* and *MYB131*.
564 In addition, Zm00001d008178, which has a homolog in rice that is an MDR-like ABC
565 transporter gene, was predicted to be regulated simultaneously by MYB16, BZIP109
566 and *MYB131*. In total, 206 genes, of which 89% (182 genes) were mainly expressed in
567 the cellularization stage, were predicted to interact with *MYB131*, *MYB16* and/or
568 *BZIP109*, including nine known TFs (*MYB8*, *MYB130*, *MYB23*, *HB20*, *HB64*, *HB84*,
569 *DOF42*, *Knox1* and *GGATA12*), one unreported ERF TF (Zm00001d016535) and 18
570 seed-specific genes. The closely related community formed by these genes might be
571 important for cellularization. *ZAG2* was predicted to interact with 50 genes, including
572 *6-phosphogluconate dehydrogenase1* (*PGD1*), *adenine phosphoribosyl transferase1*
573 (*APRT1*), *aldehyde dehydrogenase2* (*ALDH2*) and *SBP-domain protein4* (*SBP4*).
574 *BZIP11* was also predicted to interact with 50 genes, including *cytochrome c*
575 *reductase1* (*CCR1*), *rotten ear3* (*RTE3*) and *Homeobox-tf 28* (*HB28*) (**Supplemental**
576 **Figure 9 and Supplemental Data Set 8**).
577

Specific genes expressed during differentiation stage (Stage IV). About half of seed-specific genes (52%, 569/1,093) identified here were highly expressed during endosperm differentiation (102~144 HAP), and can be further divided into two sub-groups (**Figure 5 and Supplemental Data Set 5**) by combining their expression patterns during the whole development stage of seed (Chen et al., 2014). There are 392 genes including 44 TFs in group IV-A, which displayed high expression in the 6 and 8 DAP endosperm but low or no expression in later endosperm (**Figure 5**). Of these genes in group IV-A, 51.8% (203/392) (**Supplemental Data Set 6**) were identified as subregion specific genes in seed according to the RNA-seq results of LCM samples collected at 8 DAP (Zhan et al., 2015). About 77% (157/203) of these are specifically expressed in defined compartments of endosperm, including well-known ESR-specific genes (*Esr1*, *Esr2*, *Esr3*, *Esr6*, *Meg14* and *Male Sterile8*), AL-specific genes (*Al9*, *WOX2b*, *Cadtfr12*, *Cadtfr14* and *Sbt1*), BETL-specific genes (*Bap2*, *Betl-3*, *9*, *10*, *Ebe2*, *Meg6*, *Meg13*, *MYBR81* and *MRP1*) (**Supplemental Data Set 6**). These results suggested that the genes in group IV-A might play an important role in endosperm cell types differentiation.

Interestingly, we found defense response related genes were overrepresented in group IV-A. For example, *Esr6* in IV-A is a defensin gene specifically expressed in ESR and plays a protective role (Balandin et al., 2005). Some of the BETL-specific genes in IV-A, including *Ebe*, *Betl3* and *Bap2* in IV-A were also suggested to involve in defense against pathogen entry into the developing seed (Magnard et al., 2003; Barrero et al., 2006). Exploring the function of these seed-specific defense-response-related genes might be helpful for understanding the establishment of defense response mechanism in new differentiated tissues in early endosperm development.

The top 5 seed-specific TFs with the most connections in group IV are *ARF17*, *MYBR81*, *MYB80*, *BZIP46* and *HB118*, which were predicted to interact with 73, 67, 53, 48 and 48 genes, respectively (**Figure 6B, Supplemental Figure 9 and**

608 **Supplemental Data Set 8**). These five TFs are all in group IV-A. As shown in Figure
609 6B, we found *MYBR81*, *HB118* and *BZIP46* and their interacting genes formed a
610 closely related community. There were 17 genes, including eight previously
611 characterized genes (*UMC1149*, *IBD7*, *GPC4*, *HEX8*, *ABI3*, *WOX2*a, *PMPM2* and
612 *EXBP6*) were predicted to be simultaneously regulated by *MYBR81*, *HB118* and
613 *BZIP46*.
614
615 In total, 97 genes were predicted to interact with *MYBR81*, *HB118* and *BZIP46*, which
616 were all mainly expressed in the differentiation stage. About half of these genes (48%,
617 47/97) were seed-specific genes, including 11 seed-specific TFs (*LBD7*, *BZR3*,
618 *MYBR29*, *MRP1*, *BHLH167*, *GATA8*, *MYB155*, *MYB83*, *WOX2a* and two unreported
619 TFs). Notably, *MYBR81* and *HB118* are two BETL specific expression TFs (Zhan et
620 al., 2015). Moreover, we found 13 of their interacting genes were identified as
621 BETL-expressed genes, including five annotated genes (*HB96*, *MYBR29*, *MRP1*,
622 *EXPB7* and *HEX8*). Of which *MRP1*, an important BETL regulator (Gomez et al.,
623 2009) and *MYBR29* were predicted to be simultaneously regulated by *MYBR81* and
624 *HB118*. In summary, these results suggested the important role of TFs like *MYBR81*,
625 BZIP46 and *HB118* in endosperm differentiation, especially in the BETL
626 differentiation.
627
628 Compared to genes in group IV-A, genes in group IV-B (177 genes including 16 TFs)
629 were continuously expressed from initial differentiation to endosperm maturation
630 (**Figure 5 and Supplemental Data Set 5**), which suggested that genes in group IV-B
631 might be mainly involved in specific biological processes (such as grain filling) in
632 well differentiated endosperm that happen over a prolonged period of time. For
633 example, *ZmSWEET4C* in IV-B encodes a hexose transporter that transfers cell wall
634 invertase–derived hexoses into or across the BETL, a key step in seed filling. Indeed,
635 seed filling is defective for the mutants of both *ZmSWEET4c* and its rice ortholog
636 *OsSWEET4*, with a strong empty pericarp phenotype (Sosso et al., 2015). *Floury3*
637 (*FL3*) in IV-B is a maternally expressed imprinted gene, which encodes a PLATZ

(plant AT-rich sequence and zinc binding) family protein (Li et al., 2017). *FL3* can modulate the biogenesis of tRNAs and 5S rRNA through interactions with RNA polymerase III transcription machinery, which may underlie endosperm development and storage reserve filling (Li et al., 2017). The semidominant negative mutant of *fl3* exhibits severe defects in the endosperm (Li et al., 2017). *Tryptophan aminotransferase related1* (*ZmTar1*) and *ZmYUC1*, two genes involved in different tryptophan-dependent pathways of IAA biosynthesis, were also found in IV-B. *ZmTar1* is an endosperm-specific paternally expressed imprinted gene critical for the indole-3-pyruvic acid (IPA) branch (Chourey et al., 2010; Zhang et al., 2011). *ZmYUC1* is an endosperm-specific IAA biosynthesis protein gene critical for the tryptamine (TAM) branch (Chourey et al., 2010). In *zmyuc1* mutant, free IAA level is reduced and has approximately 40% less dry mass as compared with wild-type (Bernardi et al., 2012). Both ZmTar1 and ZmYUC1 are important for highly complex homeostatic control of IAA levels in maize endosperm (Chourey et al., 2010).

There were 13 annotated TFs (*BHLH167*, *MYB155*, *NRP1*, *WOX2a*, *MYB83*, *GATA8*, *DOF36*, *MYB127*, *NAC130*, *SCL1*, *EREB137*, *EREB167* and *GATA33*) and three unknown TFs (Zm00001d006319, Zm00001d040952 and Zm00001d017899) identified in IV-B. The two TFs with the most connections in group IV-B are *BHLH167* and *MYB155*, which were both predicted to interact with 40 genes. The *BHLH167* was very recently reported as *Opaque11* (*O11*) (Feng et al., 2018). O11 is a major regulator of maize endosperm metabolism, development, and stress response, which regulates nutrient metabolism through directly regulating carbohydrate metabolic enzymes and the upstream regulators, including O2 and PBF (Feng et al., 2018). The starch and protein accumulation were decreased in *o11*, a classic seed mutant with a small and opaque endosperm (Nelson, 1981). The *MYB155* is reported to be highly expressed in the maize endosperm and involved in the process of starch biosynthesis (Xiao et al., 2017). Collectively, these identified seed-specific genes and TFs in group IV-B might be critical for grain filling process, according to the function of the known genes and TFs in this group.

Specific genes expressed during more than one of the four stages. In total, 230 seed-specific genes, including 24 TFs, were expressed at more than one of the four stages, and they can be further divided into two groups (**Figure 5 and Supplemental Data Set 5**). Genes in group 6 (187 genes, 22 TFs) displayed high expression at 0~144 HAP with low or even no expression at the other later stages of seed development (**Figure 5**). The genes in group 6 were related to cell wall organization, sexual reproduction and aspartic-type endopeptidase activity. Compared to seed-specific genes in group 6, the genes in group 7 (43 genes including 2 TFs) were found to be expressed at later stages of seed development. The overall expression levels of genes in group 7 in embryo are obviously higher than that in endosperm, implying many of these genes might be mainly involved in the specific biological processes occurred in the embryo. For example, two TFs in group 7, *viviparous-1* (*Vp1*) and *WRI1 transcription factor2* (*Wri2*), are reported to play important roles in embryo development. *Vp1* is reported to be highly expressed in maize embryo and controls the anthocyanin pathway by regulating *colored aleurone1* (*C1*). The embryo of *vp1* mutant displays reduced sensitivity to the hormone abscisic acid, resulting in precocious germination, and blocked anthocyanin synthesis in aleurone and embryo tissues (McCarty et al., 1989). *Wri2* is a key regulator of seed oil biosynthesis in maize. It showed a strong transcriptional induction during the early filling stage of the embryo in maize and could complement the reduced fatty acid content of Arabidopsis *wri1-4* seed (Pouvreau et al., 2011). In summary, the seed-specific genes identified here will be very helpful to understand the specific biological processes occurred during seed development, especially for that of early stages.

DISCUSSION

In this study, we constructed a high-temporal-resolution dynamic transcriptome landscape of early maize seed development by sampling 31 time points from 0~144 HAP at intervals of 4 hours (0~72 HAP) or 6 hours (72~144 HAP). Our tissue samples for transcriptomic analysis contained the nucellus and embryo sac. The

nucellus will be degraded gradually after double fertilization (Greenwood et al., 2005; Russell, 1979), while the embryo sac is the area of the initiation of embryo and endosperm development (Chaudhury et al., 2001; Dumas and Mogensen, 1993). As shown by previous morphological observation (Leroux et al., 2014), the major part of our samples is the nucellus, which should make the largest contribution to the transcriptome data that we generated. However, as the transcriptome reprograming is extremely active in the embryo sac during early seed development (early embryo and endosperm), even though nucellus tissue constitutes a big part of our samples, the dynamic we observed could mostly reflect the activity of earlier endosperm and embryo, particularly for the endosperm tissues which enlarged much more than the embryo in the earlier stages.

The dynamic transcriptome data provided here clearly demonstrated the four key development stages within the early seed, including the stage around double fertilization, coenocyte formation, as well as cellularization and differentiation of endosperm, which the occurrence times revealed here are consistent with those reported previously (Chen et al., 2017; Sabelli and Larkins, 2009; Leroux et al., 2014; Olsen, 2001). We found there are 4,453, 1,285, 2,569 and 3,614 genes mainly expressed at the stages of around double fertilization, coenocyte formation, cellularization and differentiation, respectively (**Table 1**), during the early development of maize seed. This large collection of genes provides a rich resource for future functional studies, which will greatly enhance our understanding of the genetic control of early seed development. In particular, we detected 1,093 seed-specific genes, including 110 TFs, which will no doubt be the targets of future functional genomics studies. For example, through the GRN analysis, our results suggested that the seed-specific TFs *MYB131*, *MYB16* and *BZIP109* might be critical for endosperm cellularization, and *MYBR81*, *BZIP46* and *HB118* might play a key role in endosperm differentiation, which together with their predicted target genes formed a closely related community at cellularization and differentiation stages, respectively. Nevertheless, the exact roles of these seed-specific genes in early seed development

remain to be determined.

In summary, our data set provides a high-temporal-resolution atlas of gene expression during early maize seed development. This data provides a much-needed high resolution gene expression profiling during all the stages of early seed development. The seed-specific genes (stage specific genes) and particularly the TF-target genes GRN uncovered here provide a solid foundation for the identification of key players involved in determining each specific cell types of the early seed development in the future.

METHODS

Plant material collection and RNA sequencing

The maize (*Zea mays*) inbred line B73 was grown in the field in May of 2016 in Beijing, China, and was pollinated in July. All the individual plants were self-pollinated at the same time. The nucellus (embryo sac included) was collected by manual dissection, frozen immediately in liquid nitrogen and stored at -80℃ before processing. Two biological replicates were set up for each time points. Each replicate was obtained by pooling samples from at least three plants.

Total RNA was extracted using TRIzol reagent (Invitrogen). RNA-seq libraries were constructed according to the manufacturer's protocol of Vazyme mRNA-seq library preparation kit (Vazyme) and were sequenced to generate 150-nucleotide paired-end reads on an Illumina HiSeq platform.

Read mapping and analysis

The B73 reference genome (RefGen_v4) (Jiao et al., 2017) was downloaded from http://ensembl.gramene.org/Zea_mays/Info/Index. After removing low quality reads using the SolexaQA (V2.5) software (Cox et al., 2010), Illumina sequencing reads were mapped to the B73 reference genome using Hisat2-2.0.4 (Kim et al., 2015) with

default settings for parameters. The bam files of uniquely mapped reads were used as inputs for the Cufflinks (V2.2.0) software (Ghosh and Chan, 2016) and FPKM (fragments per kilobase of transcript per million fragments) values were calculated to measure the expression levels of genes. We calculated the Pearson correlation coefficient between biological replicates with the normalized expression levels of $\log_2$ (FPKM value + 1).

Hierarchical clustering was performed by the MeV (V4.9) software (http://www.tm4.org/mev.html) with the HCL method. PCA was performed using the prcomp function in R software (R, 2013) with default settings to facilitate graphical interpretation of relatedness among 31 different time points samples. The transformed and normalized gene expression values with $\log_2$ (FPKM + 1) were used for hierarchical clustering, and the Z scores of the genes were used for the analysis of PCA.

Gene coexpression and functional enrichment analysis

The MeV (V4.9) software with the k-means method was used for coexpression analysis for 31 different time points samples. The normalized expression values of genes, which were calculated by dividing their expression level at different time points with their maximum observed FPKM. The figure of merit (Yeung et al., 2001) was used to determine the optimal cluster number. Functional category enrichment for each coexpression module was evaluated with the MapMan (v3.6.0) functional annotation (Thimm et al., 2004). Before conducting the MapMan annotation, we choose the longest protein of each gene as a representative protein and run the Mercator with default settings. Fisher's exact test was used to examine whether the functional categories were overrepresented for a given module. Resulting P values were adjusted to Q values by the Benjamini-Hochberg correction, and a false discovery rate of 5% was applied.

Identification of seed-specific gene expression

31 different time points seed samples collected here and 23 non-seed RNA-seq data

(Bolduc et al., 2012; Davidson et al., 2011; Jia et al., 2009; Li et al., 2010; Wang et al., 2009) downloaded from the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/) were used for identification of seed-specific genes. The method we described previously was used (Chen et al., 2014). Briefly, the expression levels across all of the samples were normalized using $\log_2$ (FPKM + 0.01). Then, we calculated Z scores of the given gene in different seed samples compared with the non-seed samples using the normalized expression level. The gene was determined to be seed specifically expressed if it had a Z-score above 3 in at least one of the seed samples. Then, combined with the transcriptome data we generated previously (Chen et al., 2014), we further explored the expression patterns of seed-specific genes we identified in later stage of embryo and endosperm development by performing coexpression analysis using the MeV (V4.9) software.

The subregion specific genes mentioned in this paper were identified based on their compartment specificity (CS) scores in different subregions of seed reported previously (Zhan et al., 2015). A gene was defined as subregion specific gene if its compartment specificity (CS) score is larger than 0.5.

Gene regulatory network inference

We used context likelihood of relatedness (CLR) algorithm method (Faith et al., 2007) to construct TF-related gene regulatory network. Mutual information (MI) for calculating the expression similarity between the expression levels of TF and gene pairs were calculated by R software (entropy package) (R, 2013). The CLR calculated regulation strength by comparing the MI between a TF and its gene pairs to the background network distribution of MI for all TFs and gene pairs that include one of the TFs and its target. The final formula is $f(Z_i, Zj) = \mathrm{SQRT}(Z_i^2+Z_j^2)$, where Z_i^2 is the Zscore between gene i and its background genes; Z_j is the Zscore between gene j and its background genes (Faith et al., 2007). Finally, we set $f(Z_i, Z_j)$ above 4.5 to identity the tightly regulation relationship between all pairs of genes and TFs.

Accession Numbers

The generated raw reads have been uploaded to NCBI's SRA database and are available under the accession number PRJNA505095. RNA-seq data as FPKM values is available via the eFP Browser engine (http://bar.utoronto.ca/efp_maize/cgi-bin/efpWeb.cgi?dataSource=Early_Seed), which 'paints' the expression data onto images representing the samples used to generate the RNA-seq data.

Sequence data for the genes mentioned in this article can be obtained from the literature based on the gene list in Supplemental Table 2.

SUPPLEMENTAL DATA

Supplemental Figure 1. Sketch of the sampled region.

Supplemental Figure 2: Validation of RNA-seq data with known genes.

Supplemental Figure 3: MapMan functional categories enriched in different coexpression modules of nucellus.

Supplemental Figure 4: Heat map of the expression patterns of genes in module I-D.

Supplemental Figure 5: Expression patterns of seed-specific genes in all nucellus and non-seed samples.

Supplemental Figure 6: Percentage of genes, tissue-specific genes, TFs, and tissue-specific TFs detected in each time point in the nucellus.

Supplemental Figure 7: Frequency distribution of the degree of nodes in the gene regulatory network.

Supplemental Figure 8: MapMan functional categories enriched in different coexpression modules of specific genes.

Supplemental Figure 9: Network hubs regulating genes in different stages.

Supplemental Table 1: Summary of RNA-Seq read mapping results.

Supplemental Table 2. Accession numbers of genes mentioned.

Supplemental Data Set 1. Expression level of genes in different samples.

Supplemental Data Set 2. List of TFs expressed in nucellus samples.

Supplemental Data Set 3. List of genes in module I-D expressed in gametes, zygotes

and early two-celled pro-embryo cells in maize.

Supplemental Data Set 4. Expression level of histone protein genes in each time point of nucellus.

Supplemental Data Set 5. Summary of seed-specific genes.

Supplemental Data Set 6. Detail information of genes in cluster IV-A and IV-B.

Supplemental Data Set 7. All 31,256 edges of the GRN.

Supplemental Data Set 8. Detail information of top 5 highly connected network specific TFs in corresponding stages I ~ IV.

AUTHOR CONTRIBUTIONS

J.L., F.Y. and W.G. designed the experiments. F.Y, W.G, N.S., X.G, X.M, H.Z. and W.S. performed the experiments. F.Y., X.Z., Y.Z., and J.C. analyzed the data. E.E., A.P., and N.P. contributed to the RNA-seq data accessibility via the eFP Browser engine, F.Y., W.G. and J.L. wrote the paper. Correspondence and requests for materials should be addressed to J.L. (jlai@cau.edu.cn).

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Declaration of Interests

No conflict of interest declared.

872 **Table 1. The number of coexpressed genes and seed-specific genes detected in**
873 **different development stages of early maize seed.**
874

Developmental stage	Number of Genes/TFs	Number of Specific genes/TFs
Around double fertilization (0~16 HAP)	4,453/414	160/18
Coenocyte (20~44 HAP)	1,285/53	22/1
Cellularization (48~96 HAP)	2,569/125	112/7
Differentiation (102~144 HAP)	3,614/224	569/60
Other*	10,869/599	230/24
Total	22,790/1,415	1,093/110

875 *: The genes were expressed at more than one of the four stages.
876

Figure legends

Figure 1. Changes in the maize nucellus and seed from 0 to 144 hours after pollination (HAP).

The nucellus (included embryo sac) samples from 31 different time points were used for transcriptome analysis.

Figure 2. Transcriptome relationships among 31 time points of early maize seed development.

A, Cluster dendrogram showing four distinct development stages: around double fertilization, coenocyte, cellularization and differentiation. B, PCA of the transcriptomes of the 31 time point samples. C, Graphic representation of the embryo sac in the four distinct development stages of seed. The pollen tube is shown in orange, sperm nuclei are shown in dark blue, polar nuclei and endosperm nuclei are shown in red, the egg cell and embryo cell are shown in yellow, the basal endosperm transfer layer (BETL cell), aleurone (AL) cell and embryo-surrounding region (ESR) cell are shown in light blue, purplish red and green, respectively. D~G, The marker genes mainly expressed in the stages of around double fertilization (D), coenocyte (E), cellularization (F) and differentiation (G). The time points belong to the stage of around double fertilization, coenocyte, cellularization and differentiation are shown in light yellow, blue, green and deep yellow, respectively.

Figure 3. Gene expression pattern and functional transition over the time course.

A, Expression patterns of genes in different coexpression modules. For each gene, the FPKM value normalized by the maximum value of all FPKM values of the gene over all time points is shown. The number of genes and TFs in each module are showed on the right. B, MapMan functional categories enriched in different coexpression modules. Only significant categories (FDR < 0.05) are displayed.

Figure 4. Expression pattern of major histone protein genes in 31 time points

samples.

Figure 5. Expression patterns of seed-specific genes.

Analysis of the expression patterns of seed-specific based on the RNA-seq data of nucellus generated in this study, and the RNA-seq data of embryo and endosperm generated previously (Chen et al., 2014). For each gene, the FPKM value normalized by the maximum value of all FPKM values of the gene over all the samples used for analysis. The number of genes and TFs in each group are showed on the left.

Figure 6. Network hubs regulating genes in different seed development stages.

A, Network hubs (*MYB131*, *MYB16* and *BZIP109*) regulating genes in cellularization stage. B, Network hubs (*BZIP46*, *MYBR81* and *HB118*) regulating genes in differentiation stage. Color codes indicate that the gene displayed with the peak expression in the corresponding stages. Yellow, around double fertilization; Gray, coenocyte; light green, cellularization; Pink, differentiation. Light blue, the genes expressed at more than one of the four stages. Genes are shown as small circles, seed-specific genes are shown as big circles, non-seed specific TFs are shown as small triangles, seed-specific TFs are shown as big triangles.

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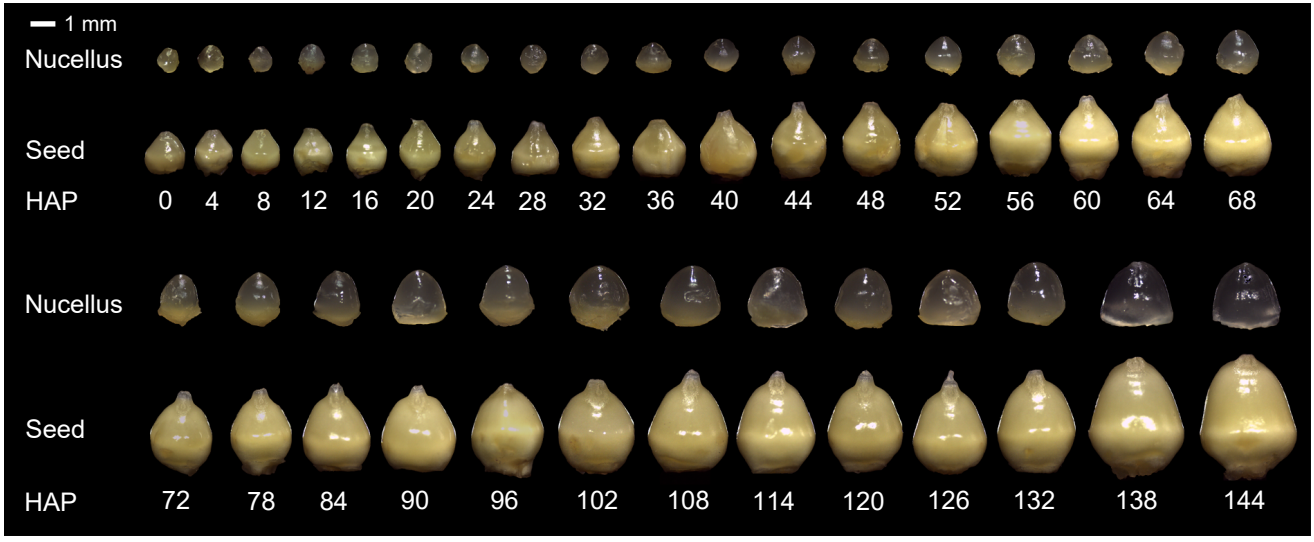

Figure 1. Changes in the maize nucellus and seed from 0 to 144 hour after pollination (HAP).
The nucellus (included embryo sac) samples from 31 different time points were used for transcriptome analysis.

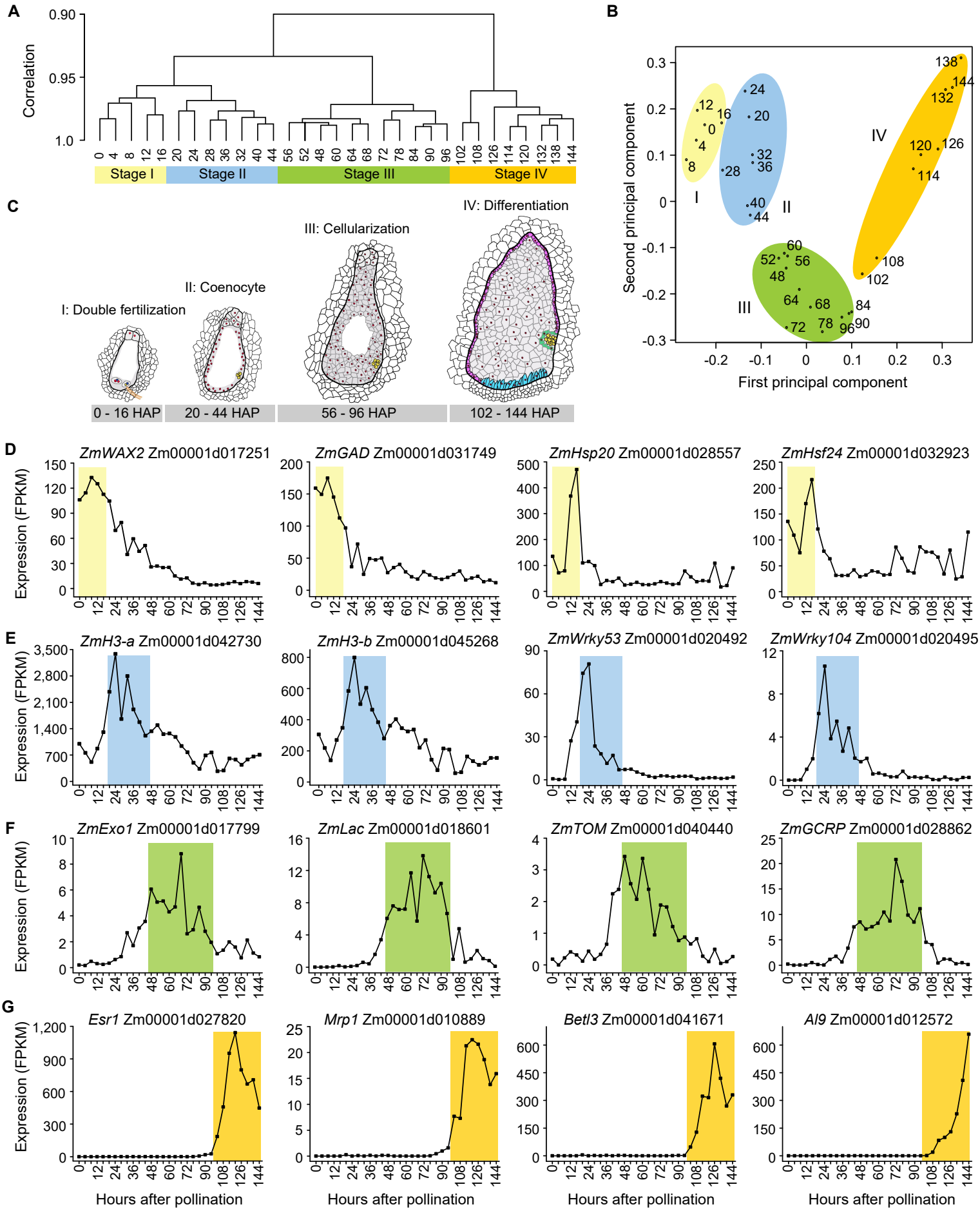

Figure 2. Transcriptome relationships among 31 time points of early maize seed development.

A, Cluster dendrogram showing four distinct development stages: around double fertilization, coenocyte, cellularization and differentiation. B, PCA of the transcriptomes of the 31 time point samples. C, Graphic representation of the embryo sac in the four distinct development stages of seed. The pollen tube is shown in orange, sperm nuclei are shown in dark blue, polar nuclei and endosperm nuclei are shown in red, the egg cell and embryo cell are shown in yellow, the basal endosperm transfer layer (BETL cell), aleurone (AL) cell and embryo-surrounding region (ESR) cell are shown in light blue, purplish red and green, respectively. D~G, The marker genes mainly expressed in the stages of around double fertilization (D), coenocyte (E), cellularization (F) and differentiation (G). The time points belong to the stage of around double fertilization, coenocyte, cellularization and differentiation are shown in light yellow, blue, green and deep yellow, respectively.

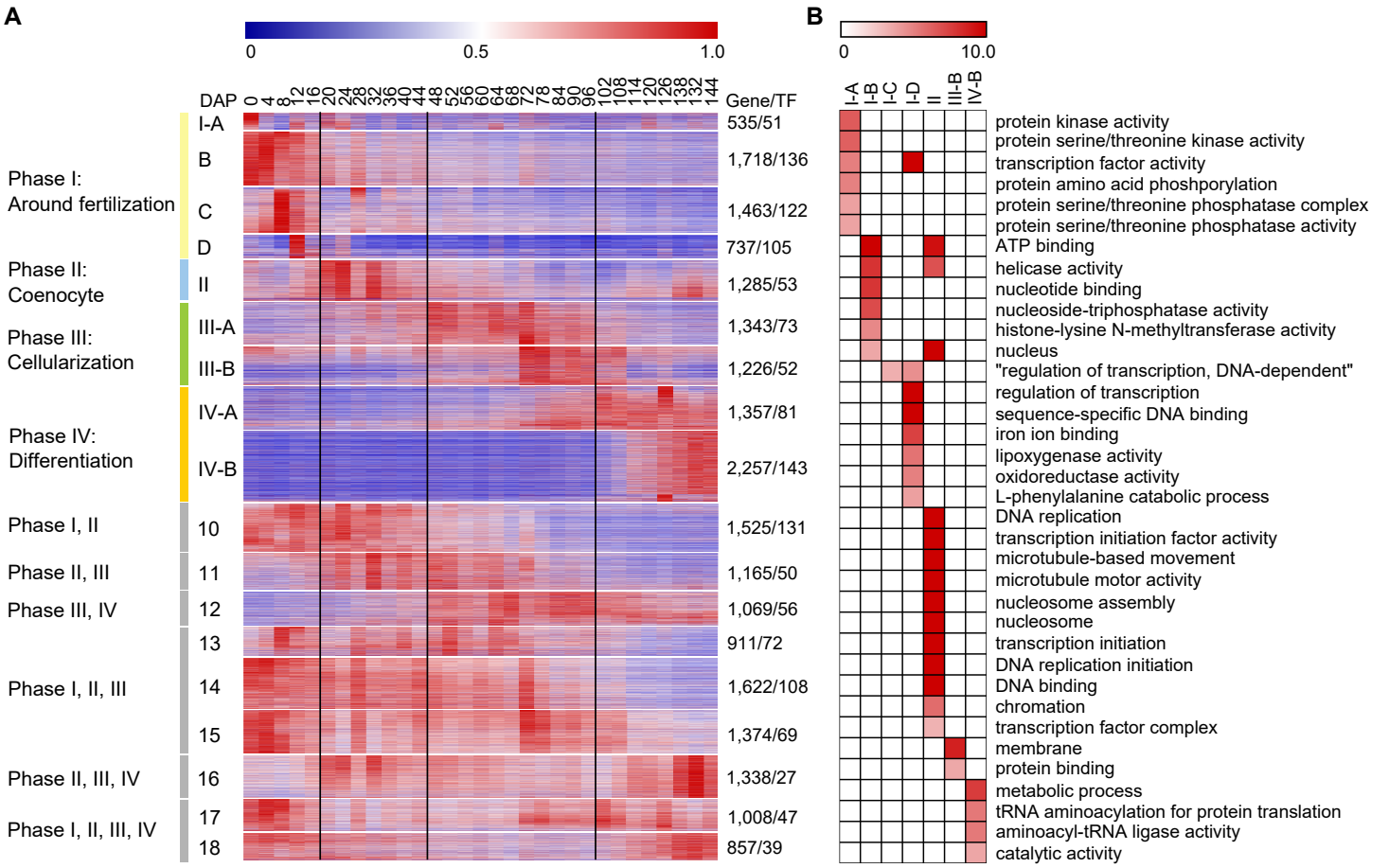

Figure 3. Gene expression pattern and functional transition over the time course.

A, Expression patterns of genes in different coexpression modules. For each gene, the FPKM value normalized by the maximum value of all FPKM values of the gene over all time points is shown. The number of genes and TFs in each module are showed on the right. B, MapMan functional categories enriched in different coexpression modules. Only significant categories (FDR < 0.05) are displayed.

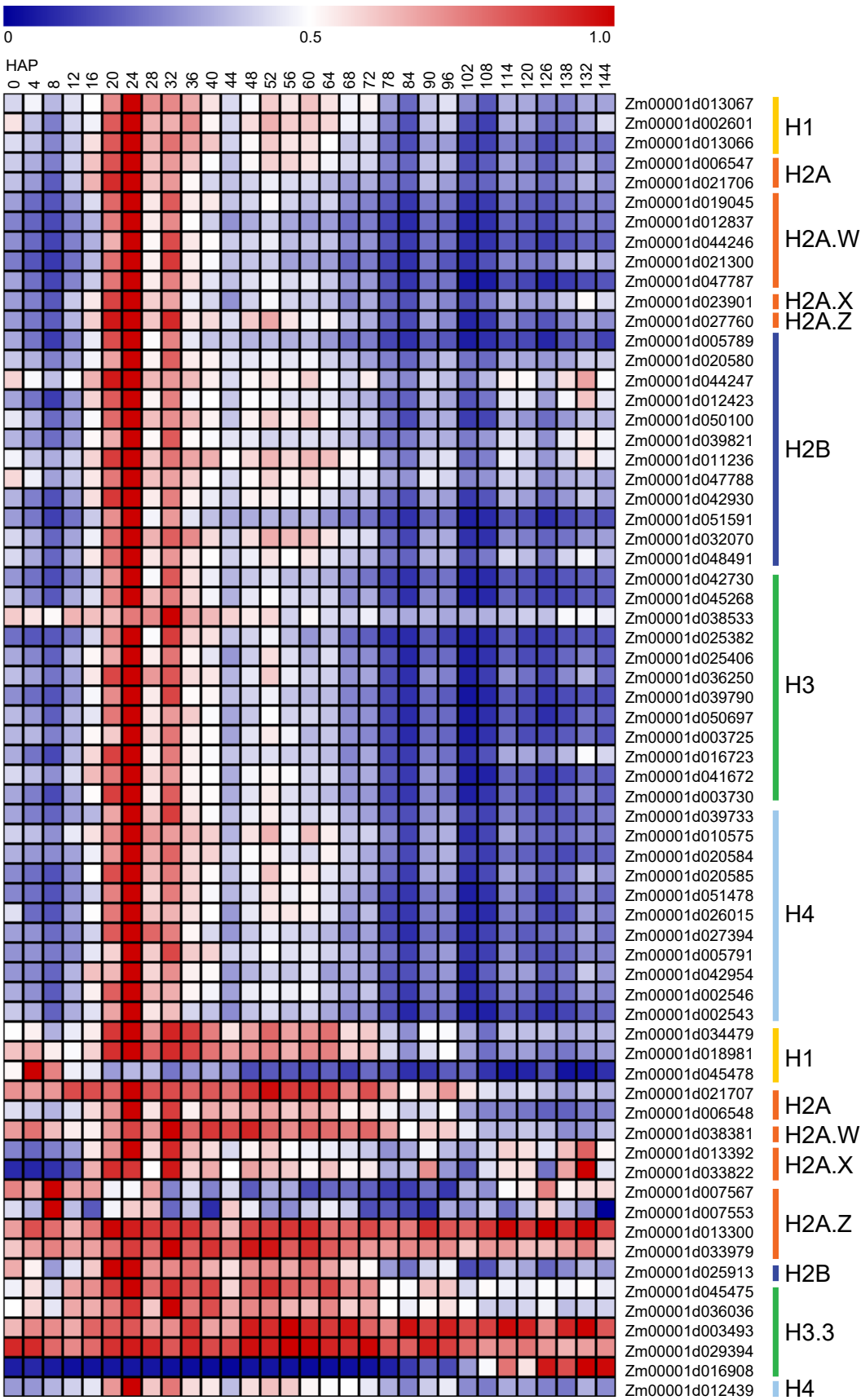

Figure 4. Expression pattern of major histone protein genes in 31 time points samples.

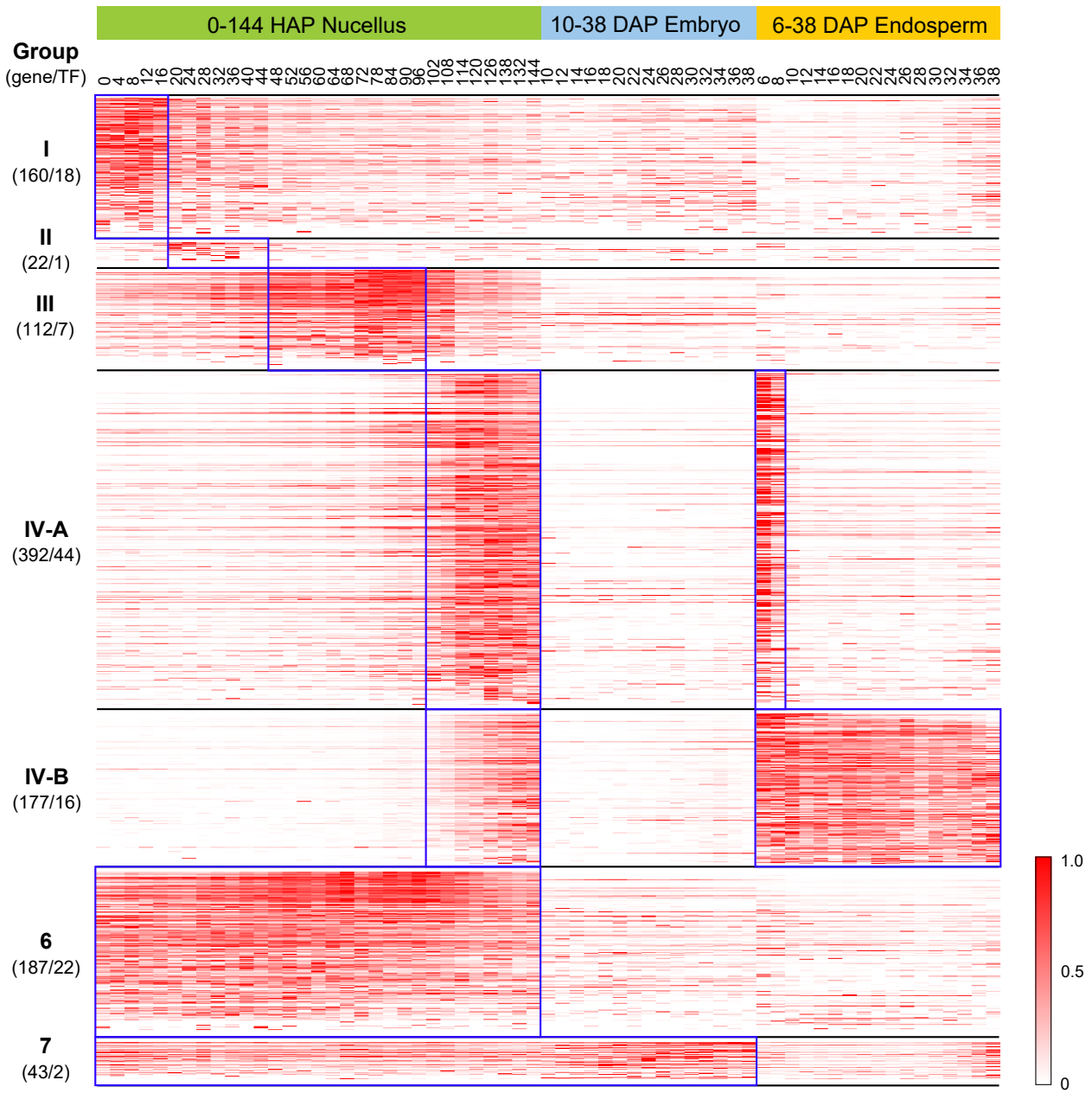

Figure 5. Expression patterns of seed-specific genes.

Analysis of the expression patterns of seed-specific based on the RNA-seq data of nucellus generated in this study, and the RNA-seq data of embryo and endosperm generated previously (Chen et al., 2014). For each gene, the FPKM value normalized by the maximum value of all FPKM values of the gene over all the samples used for analysis. The number of genes and TFs in each group are showed on the left.

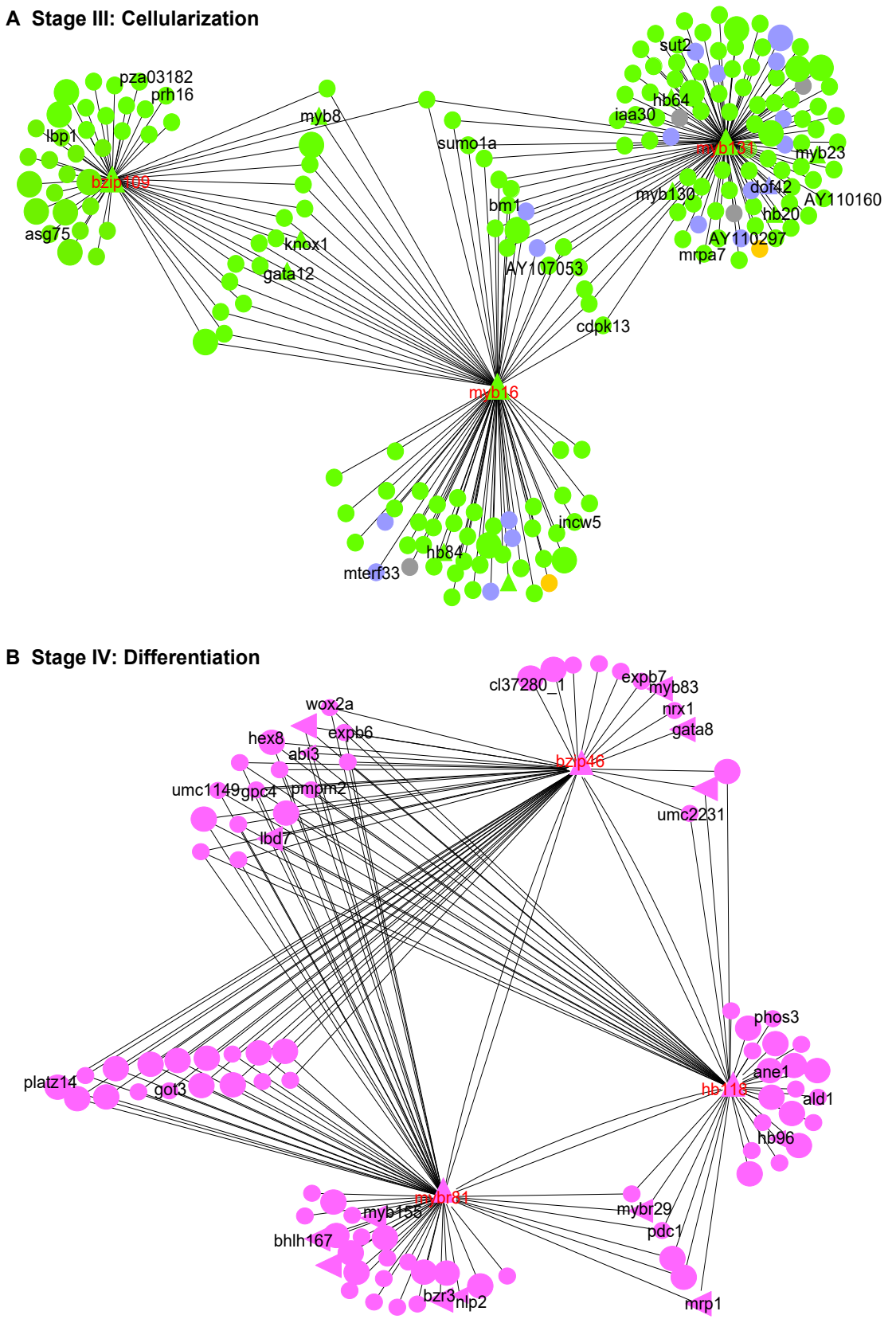

Figure 6. Network hubs regulating genes in different seed development stages.

A, Network hubs (*MYB131*, *MYB16* and *BZIP109*) regulating genes in cellularization stage. B, Network hubs (*BZIP46*, *MYBR81* and *HB118*) regulating genes in differentiation stage. Color codes indicate that the gene displayed with the peak expression in the corresponding stages. Yellow, around double fertilization; Gray, coenocyte; light green, cellularization; Pink, differentiation. Light blue, the genes expressed at more than one of the four stages. Genes are shown as small circles, seed-specific genes are shown as big circles, non-seed specific TFs are shown as small triangles, seed-specific TFs are shown as big triangles.

IN A NUTSHELL

Background: Maize seed is an important source of food, feed and biofuel materials. The early maize seed undergoes several developmental stages after double fertilization to become fully differentiated within a short period of time, but the genetic control of this highly dynamic and complex developmental processes remains largely unknown. Understanding the spatial and temporal gene expressional profile along seed development is useful for unraveling the genetic control of seed development and thus for the genetic improvement of this important crop.

Question: We wanted to know the gene activity dynamic during double fertilization, coenocyte formation, cellularization, and differentiation, four main stages of early maize seed development, especially, which genes are specifically expressed at particular stages of early maize seed development.

Findings: A total of 22,790 expressed genes including 1,415 transcription factors (TFs) were detected in early stages of maize seed development. In particular, 1,093 genes including 110 TFs were specifically expressed in the seed, most of which were newly identified in this study and displayed high temporal specificity by expressing only in particular period of early seed development. There were 160, 22, 112 and 569 seed-specific genes predominantly expressed in the first 16 hours after pollination, coenocyte formation, cellularization and differentiation stage, respectively. In addition, network analysis predicted 31,256 interactions among 1,317 TFs and 14,540 genes. The high-temporal-resolution transcriptome atlas reported here provides an important resource for future functional study to dissect the genetic control of seed development.

Next steps: We plan to select some key genes for CRISPR/Cas9-based gene editing to further explore their function in the genetic control of seed development.

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